

ENCODE COW IDENTIFICATION KIT



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ENCODE COW ID KIT
Cat. No .EIL-COW-50
Cat. No .EIL-COW-100

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1. MEAT INFORMATION

In case of meat and meat products, the animal species must be disclosed on the label. Moreover, the species authenticity can be highly relevant to consumers for economic, medical, cultural and religious reasons. Fraudulent substitution of cheaper meats in place of more expensive species, the inclusion of meat in non-meat (vegetarian) products and the presence of allergens in food products are clear examples for the importance of this issue. Several methods for species detection are based are based on identification of proteins by means of electrophoretic and/or immunological methods. However, these methods are not reliable for application in highly processed and heated products due to the protein deterioration. DNA is more stable than proteins during processing and although it can be fragmented by several processes, modern DNA methodologies like PCR based techniques still allow the identification of DNA from the species present in a sample. Real time PCR techniques are especially suitable for these products because small fragments of DNA can still be amplified and identified with high sensitivity and specificity.

2. INTENDED USE

The kit enables a qualitative detection of cow DNA in food samples by Real Time PCR, after a sample processing step. This DNA is present in the mitochondrial genome and it may be found in processed food and food industry. The test may be used with the following matrixes: cow meat, cow burger, cow milk, butter and cow cheese.

3. STORAGE CONDITIONS AND STABILITY





The kit must be stored at -20°C. The Master Mix and the Control should be stored between -20°C and 5°C. The kit must be shipped refrigerated, using coolers, and should be stored immediately on receipt. Repeated thawing and freezing should be avoided, as this may reduce the sensitivity. It should be protected from exposure to light. The kit is stable until the expiration date stated on the outer package and on the labels of the vials.

4. WARNINGS AND PRECAUTIONS

- 1. These products are exclusively for in vitro use.
- 2. Molecular Biology procedures, such as DNA extractions and PCR amplification, require qualified to prevent the risk of erroneous results, especially due to sample contamination or degradation of the nucleic acids contained in the sample.
- 3. It is strongly recommended to have dedicated areas, materials and equipment for the DNA extraction, preparation of the PCR and post-PCR procedures. Workflow in the laboratory should proceed in a unidirectional manner, from the Extraction Area to the Amplification and Detection Area.
- 4. The user should always pay attention to the following:
 - Read all the instructions provided with the product before running the assay.
 - Do not mix reagents from different batches.
 - Wear disposable gloves, laboratory coats and eye protection when handling specimens and reagents.
 - Avoid cross contamination of samples and reagent.
 - Use sterile pipette tips with filters.
 - Store and extract positive material (specimens, controls and amplicons) separately from all other reagents and add it to the reaction mix in a spatially separated facility.
 - Thaw all components thoroughly at room temperature before starting an assay. When thawed, mix the components and centrifuge.
 - Avoid contact of specimens and reagents with the skin, eyes and mucous membranes. If these solutions come into contact, rinse immediately with water and seek medical advice immediately.
 - Add the positive control after closing the all tubes.
- 5. Waste must be treated and disposed of in compliance with the appropriate safety standards.

5. ADDITIONAL MATERIAL REQUIRED

Disposable powder-free gloves

DNA extraction kit

Sterile pipette tips with filters

Tubes/Strips and accessories specific for the Real Time PCR Instrument.

6. EQUIPMENTS REQUIRED

Laminar Air Flow Cabinets/PCR Cabinets Micropipettes Microcentrifuge





Real Time PCR instrument accessories Freezer, refrigerator

7. SAMPLE COLLECTION AND TRANSPORT

This product must be used with DNA extracted from the following food samples: cow meat, cow burger, cow milk, butter and cow cheese. Samples that are to be used for DNA extraction must be collected and stored according to laboratory guidelines.

8. PROCESSING STEPS



Sample processing DNA Extraction PCR Set up and run Result and Interpretation

9. CONTENTS

Table 1:- The kit contains and Reaction Size available as below

| Reagents | 10 | 25 | 50 | 100 |
|----------------------|-----------|-----------|-----------|-----------|
| | reactions | reactions | reactions | reactions |
| Master MIX | 125 µl | 312.5 µl | 625 µl | 1250 µl |
| Cow detection mix | 25µl | 62.5 µl | 125 µl | 250µl |
| PCR grade water | 80µl | 200 µl | 400 µl | 800 µl |
| Negative control | 20 µl | 50µl | 100µl | 200 µl |
| Cow Positive control | 5 µl | 10 µl | 20 µl | 40 µl |

protected from light as excessive exposure to light may affect the fluorescent probes.

9.1 Experimental Protocol

Preparing the reaction.

Table 2:-

| Reagent | Volume per | |
|----------------------|------------------|--|
| | reaction (25 µl) | |
| Master mix | 12.5 µl | |
| Cow Detection mix | 2.5 µl | |
| Sample /Template DNA | 2 µl | |
| PCR Grade water | 8 µl | |
| Total volume | 25 µl | |





Preparing the PCR

- 1. Add 23µL of the Cow Reaction into each tube/strip.
- 2. Add 2µL of DNA sample.
- 3. Close the tubes/strips, and spin.
- 4. Place the reactions into the Real Time PCR instrument.

Program set up

| 1 Togram Set up | | | |
|-------------------------------------|------|--------|--------------|
| Stage | Temp | Time | No of cycles |
| Initial denaturation | 95°c | 15 min | 1 |
| Denaturation | 95°c | 10 sec | |
| Annealing and Extension | 60°c | 60 sec | |
| Denaturing, Annealing and Extension | | | 45 |

Passive reference dye:-ROX

- For High ROX compatible instrument dilute 5 μl ROX in 50 μl PCR Grade Water (1:10).
- For Low ROX compatible instrument dilute 0.5 μl ROX in 50 μl PCR Grade Water (1:100).

10. DATA ANALYSIS

Probe for cow detection are labeled with FAM and must be analyzed in the corresponding fluorescence channel. For each sample, compare the results obtained in each channel and interpret the results as described in the following tables:

A)Controls

To validate the assay, the controls must have the following results:

| Negative control | Positive control | Interpretation | |
|------------------|------------------|-------------------------|--|
| - | + | perfect | |
| - | - | Rerun, pipetting error | |
| + | + | Contamination, Rerun | |



^{**} If Needed, prepare a fresh dilution of ROX internal reference dye and use **0.3** µl for each reaction. The diluted ROX reference dye must be kept in a light-protected tube at 4°c



| + | - Rerun, pipetting | |
|---|--------------------|-------|
| | | error |

Note:- if the controls do not match these results, the experiment must be repeated.

B)Samples

Interpretation of sample results is summarized in the following table:

| Cow detection | IC (internal | Interpretation |
|---------------|--------------|----------------|
| FAM | control) | |

| + | + | Cow | |
|---|---|----------------------|--|
| | | detected(positive) | |
| - | + | cow not | |
| | | detected(negative) | |
| - | - | PCR inhibition, poor | |
| | | extraction, rerun | |
| + | - | PCR inhibition, | |
| | | Rerun | |

^{**}When both Cow and IC is not detected, the sample must be tested again after 1:10 dilution.

11. ADDITIONAL INFORMATION

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